

## Original Research Article

# In Vitro Susceptibility Patterns of Aqueous Extract of Green Tea against Bacteria That Cause Urinary Tract Infection in Pregnant Women Attending Shendi Hospitals

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**Article History:** | Received: 25.05.2021 | Accepted: 04.07.2021 | Published: 12.07.2021 |

**Abstract:** The aim of study is to determine the activity of aqueous extract of *Camellia Sinensis* (Green tea) on different types of bacteria isolated from 191 pregnant women attended to Shendi locality hospitals, suffering from urinary tract infection between February 2015 to February 2017 following informed consent. One hundred thirty seven bacteria were isolated, different bacteria with different Gram reactivity was isolated then identified biochemically, in vitro sensitivity testing using well diffusion technique against aqueous green tea extract was done. The largest zone of inhibition appeared in Gram positive bacteria *Enterococcus faecalis* ( $17.6 \pm 1.9$  mm), the concentration of green tea aqueous extract able to suppress the growth of bacteria was 48.6 milligram per ml against *Staphylococcus aureus* and 280.2 milligram per ml against *Escherichia coli*.

**Keywords:** *Camellia Sinensis*, Aqueous extract, Green tea, pregnant woman, Shendi, Sudan.

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## BACKGROUND

In the human females, the urinary tract has an important relation with the reproductive organs. In the non-pregnant state, the uterus lies behind and partly over the bladder while in the pregnant state the enlarging uterus effects on urinary tract (Mittal & Wing, 2005).

Urinary Tract Infections has become common hospital-acquired infection rating of about 35 % of nosocomial infections, and it is the second common cause of bacteraemia in hospitalized patients (Epoke, *et al.*, 2000).

UTI is a common problem during pregnancy (Mittal & Wing, 2005). This usually begins in first trimester and peaks during second and third trimester (Van Brummen, *et al.*, 2006). UTI can lead to serious complications if left untreated, (Mazor-Dray, *et al.*, 2009). Also it has been observed that asymptomatic bacteriuria can lead to cystitis and pyelonephritis (Barnick & Cardozo, 1991), which can lead to acute

respiratory distress, transient renal failure, sepsis and shock during pregnancy (Gilstrap & Ramin, 2001).

Screening of pregnant women for UTI can minimize it, and the associated complications (Millar & Cox, 1997). Recently various risk factors of UTI during pregnancy have been reported; perhaps these are varied according the geographical, social and biological settings (Haider, *et al.*, 2010). *Escherichia coli* with its multidrug resistant strains- has been found to be the commonest cause of UTI among pregnant women (Dalzell & Lefevre, 2000; Kariuki, *et al.*, 2007).

In recent years, the health benefits of consuming green tea, including the prevention of cancer and cardiovascular diseases, the anti-inflammatory, antiarthritic, antibacterial, antiangiogenic, antioxidative, antiviral, neuroprotective, and cholesterol-lowering effects of green tea and isolated green tea constituents are under investigation. However, adding green tea to the diet may cause other serious health concerns (Chacko, *et al.*, 2010).

The health-promoting effects of green tea are mainly attributed to its polyphenol content, particularly flavanols and flavonols, which represent 30% of fresh leaf dry weight. Recently, many of the forementioned beneficial effects of green tea were attributed to its most abundant catechin, (-)-epigallocatechin-3-gallate (EGCG). Green tea extracts are more stable than pure epigallocatechingallate, one of the major constituents of green tea, because of the presence of other antioxidant constituents in the extract.

In general, herbal medicines are complex mixtures of different compounds that often act in a synergistic fashion to exert their full beneficial effect (Chacko, *et al.*, 2010).

Tea is one of the most popular beverages consumed worldwide. Tea, from the plant *Camellia sinensis*, is consumed in different parts of the world as green, black, or Oolong tea. Among all of these, however, the most significant effects on human health have been observed with the consumption of green tea (Cabrera, *et al.*, 2006). The association between tea consumption, especially green tea, and human health has long been appreciated (Weisburger, 2000; Sato & Miyata, 2000). Some studies have suggested an inverse association between green tea consumption and the risk of kidney stone formation (McKay & Blumberg, 2002; Ishizuk, *et al.*, 2003).

A series of good conducted studies show spatially in Japan that tea extracts have several useful antibacterial effects that can inhibit and kill *Staphylococcus aureus*, *Staphylococcus epidermidis* which are urinary pathogens, *Salmonella typhi*, *Salmonella typhimurium*, *Salmonella enteritidis*, *Shigella flexneri*, *Shigella dysenteriae*, and *Vibrio* spp, including *Vibrio cholera* (Hamilton-Miller, 1995).

## MATERIALS AND METHOD

### Study design

Hospital based cross sectional study.

### Study duration

From February 2015 to February 2017

### Study population

Pregnant females in Shendi hospitals - Sudan

### Sample size

Sample was calculated from free online web site <http://www.calculator.net> by using confidence level 95% and confidence interval 8%, One hundred ninety one.

$$\text{Sample size} = Z^2 \times (p) \times (p-1) / C^2$$

Z = Z value – confidence level 95% (1.96),

C = Confidence interval (.08 = ± 8).

P = standard division (0.5).

The calculated sample size based on this formula was 191.

### Ethical approval

The proposal was reviewed and approved ethically by the scientific and the ethical committee of Shendi University.

## DATA COLLECTION

Data was collected by using questionnaire.

### Study setting

Shendi locality, River Nile State, Sudan. Shendi is a town in northern of Sudan beside the River Nile and Almatamah town also far 150 km northeast from Khartoum the capital of Sudan (16°41'N 33°25'E). The area is inhabited by the Ga'aleen Tribe.

### Specimen collection

The patient was given a sterile non-opened, wide mouth, leak proof container and requested by the lab to collect at least 10 ml of urine specimen. The container was labeled with the date, the name and number of the patient, and the time of collection. When immediate delivery to the laboratory was not possible, the patient was requested to refrigerate the urine at 4–6 °C until delivery not more than 24 hours.

### Culture of urine specimen

Urine sample were mixed well by rotating urine container several times. Beside opened Bunsen burner urine container was opened and Nichrome loop was inserted after sterilization by flaming and cooling. Small amount of urine sample was taken by loop and inoculated by making firstly well in Cysteine lactose electrolyte deficient agar (CLED) media then making primary lines from the well then secondary lines from primary lines then tertiary lines from secondary lines finally zigzag from last line of tertiary lines. The inoculated plates were incubated in incubator at 37°C for 24h under aerobic condition.

### Interpretation of culture growth

The plates were observed for any bacterial colonies grow significantly. The bacteria were well isolated and then identified by colonial morphology, Gram stain and biochemical tests.

### Preparation of plant extract

The plant extracts were prepared using the solvent water. 15g of green tea dry leaves (Vietnam green tea) minced to powder were taken and homogenized with 100 ml of the distilled water (Kumar, *et al.*, 2012). The mixture was left in hot air oven under 80°C for 2 hours (Uzunalic, *et al.*, 2006). After that left to cool and filter the mixture by piece of gauze then by filter paper of Whatman No.1 manufacture. The extract was heated in hot air oven under 60°C to make stock crude material. The crude was weight by sensitive balance and 0.5g then dissolved in 1ml of sterile distilled

water to obtain concentration 500mg/ml then diluted in tubes by using two fold dilution to make concentration 250,125,62 and 31 mg/ml.

**Procedure of inoculation in Mueller Hinton agar plates and applying green tea extract**

By the loop 3–5 colonies were taken and also must be similar in appearance, then transfer the colonies tube containing sterile normal saline and mixed then compared the tube with the standard turbidity and adjusted the turbidity of the test suspension to that of the standard by adding more bacterial colonies or more sterile normal saline. The plates were inoculated by dipping a sterile swab into the tube containing bacterial suspension.

The excess of bacterial suspension was removed by gentle pressing and rotating the swab against the sides of the tube above the level of the suspension. The swab was masked all over the surface of the plate media with rotating each time to ensure complete covering. Finally. The plate was left to dry for a 1- 2 minutes at room temperature with the lid closed. By using glass porer of size 6 mm in diameter, 5 pores were made in agar plate then the pores were filled by green tea extract by using automatic pipette in volume 50 µ.l of concentrations 500mg/ml (Stock concentration), 250 mg/ml, 125 mg/ml, 62.5 mg/ml and 31 mg/ml.The plates were incubated for 24h in incubator under aerobic condition in 37°C.

**Interpreting the sensitivity of green tea extract**

The diameter of each inhibited growth zone had been measured by graduated ruler under-surface of the plate without opening the cover and recorded in mm.

**DATA ANALYSIS**

Data was analyzed by using online web site <https://www.graphpad.com> to determine the mean and standard deviation of the data. Proportional data were presented as frequencies and percentages.

**RESULT**

A total of one hundred ninety one pregnant women with UTI were contributed in this study. The age mean was (25.4 ± 6.7) years. Most of them were within the age group less than 24 and 25 – 34 years (47.7%, 47.1%) respectively (Table 1).More than 50% were in their 3d trimester (Table 2). One hundred forty six (76%) of the pregnant women drinking tea while they were pregnant and 145 (99.3%) drink black tea showed in (Table 3). The main causative agent of UTI in the study population was *S. aureus* (24.8%), followed by *E. coli* (21.9%) and the least causative agents were *Hafnia alvei* and *Proteus vulgaris* (0.7 %) (Table 4). The Gram negative bacilli isolated were *E. coli* represented the main causative agent between Gram

negative bacilli bacteria 30 (50) and lowest causative Gram negative bacilli were *Proteus vulgaris* and *Hafnia alvei* 1 (1.6%) (Table 5). The Gram positive cocci isolated was *S. aureus* 34 (44.2%) and it is the main causative agent between Gram positive cocci bacteria and lowest causative Gram positive cocci was *Enterococcus fecalis* 5 (6.5 %) (Table 6). In 1<sup>st</sup> trimester *Citrobacter freundii* was the main causative agent of UTI 5 (29.4%), *E. coli* and *Klebsiella pneumoniae* were least isolated agents 1 (5.9%).In 2<sup>nd</sup> trimester *S. aureus* was the main causative agent of UTI 10 (27.1%), *Enterobacter cloacae*, *S. saprophyticus* and

*E. fecalis* were least isolated agents 1 (2.7%).In 3<sup>d</sup> trimester also *S. aureus* was the causative agent of UTI 21 (25.3%), *Proteus vulgaris*, *Hafnia alvaei*, *Citrobacter freundii* and *Enterobacter cloacae* were least isolated agents1 (1.2%) (Table 7). Antimicrobial susceptibility of green tea extract has largest zone of inhibition against *Enterococcus fecalis* was (17.6 ± 1.9 mm) and least zone of inhibition against *Proteus vulgaris* and *Enterobacter cloacae* was (8 ± 0 mm) (Table 8 ).The results of antimicrobial activity of extract was compared with Standard drugs (Gentamicin) as positive control. Antibacterial activities of tea extract were tested by well diffusion method. The concentrations of green tea water extract used was 500, 250, 125, 62.5 and 31 mg/ml. MIC results of water extracts of green tea on test organisms which the lowest concentration of green tea extract able to suppress the growth of bacteria was (48.6 mg/ml) appear against *S.aureus* followed by *Staphylococcus saprophyticus* (58 mg/ml), *Enterococcus fecalis* (62.8 mg/ml), *Micrococcus luteus* (66.8 mg/ml) *Staphylococcus epidermidis* (68.2 mg/ml), *Hafnia alvei* (250 mg/ml), *E. coli* (280.2 mg/ml), *Citrobacter freundii* (312.5 mg/ml), *Citrobacter diversus* (325 mg/ml), *Enterobacter cloacae*, *Klebsiella pneumoniae* and *Proteus vulgaris* (500mg/ml) (Figure 1).

**Table-1: Show population distribution according to ages**

Ages groups	Number	%
Less than 24	91	47.7%
25 – 34	90	47.1%
35 – 44	9	4.7%
More than 44	1	0.5%
<b>Total</b>	<b>191</b>	<b>100%</b>

Age (mean ± SD) = (25.4 ± 6.7)

**Table-2: Show distribution of the population according to trimester**

Trimester	Number	%
First	24	12.6%
Second	60	31.4%
Third	107	56.0%
<b>Total</b>	<b>191</b>	<b>100</b>

**Table-3: Show distribution of population according to tea type used**

Type of tea used	Number	%
Black	145	99.3%
Green	1	0.7%
<b>Total</b>	<b>146</b>	<b>100%</b>

**Table-4: Show percentage of bacteria isolated from urine of study population**

Bacteria name	Number	%
Citrobacter diversus	10	7.30%
Citrobacter freundii	6	4.40%
Escherchia coli	30	21.9%
Enterobacter cloacae.	2	1.50%
Enterococcis fecalis	5	3.60%
Hafnia alvei	1	0.70%
Klebsiella pneumoniae	10	7.30%
Micrococcus luteus	15	11%
Proteus vulgaris.	1	0.70%
Staphylococcus aureus	34	24.8%
Staphylococcus epidermidis	16	11.70%
Staphylococcus saprophyticus	7	5.10%
<b>Total</b>	<b>137</b>	<b>100</b>

**Table-5: Show Gram negative Bacteria isolated from urine of study population**

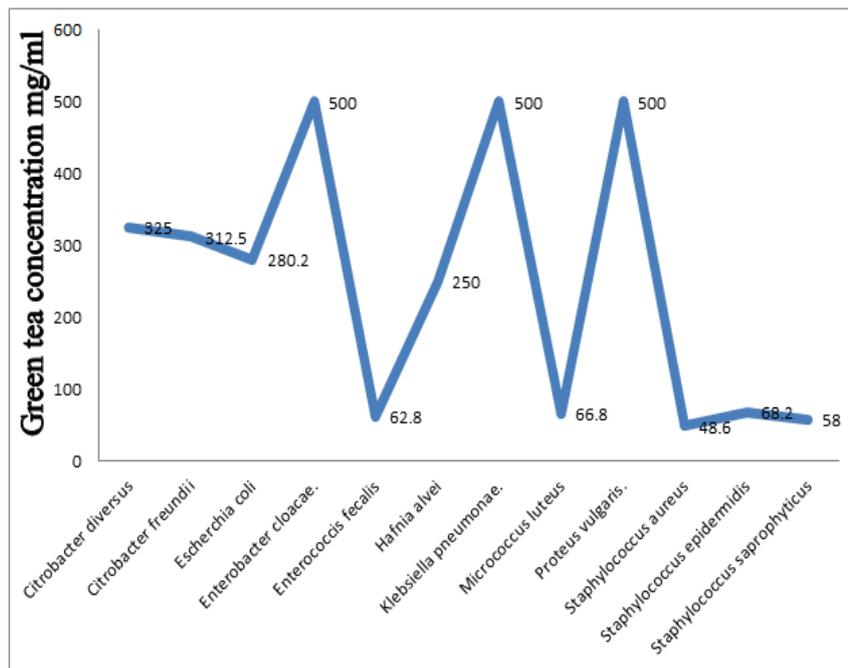
Bacteria isolated	Number	%
Citrobacter diversus	10	16.7%
Citrobacter freundii	6	10%
Escherchia coli	30	50%
Enterobacter cloacae.	2	3.4%
Hafnia alvei	1	1.6%
Klebsiella pneumonia	10	16.7%
Proteus vulgaris	1	1.6%
<b>Total</b>	<b>60</b>	<b>100%</b>

**Table-6: Show Gram positive Bacteria isolated from urine of study population**

Bacteria isolated	Number	%
Staphylococcus aureus	34	44.2%
Staphylococcus epidermidis	16	20.7%
Staphylococcus saprophyticus	7	9.1%
Enterococcus fecalis	5	6.5%
Micrococcus luteus	15	19.5%
<b>Total</b>	<b>70</b>	<b>100%</b>

**Table-7: Show percentage of isolated bacteria from urine of pregnant woman according to trimester**

Bacteria name	Total No.	1 <sup>st</sup> trimester		2 <sup>nd</sup> trimester		3 <sup>d</sup> trimester	
		n	%	n	%	n	%
Citrobacter diversus	10	0	0%	4	10.8%	6	7.2%
Citrobacter freundii	6	5	29.4%	0	0%	1	1.2%
Escherchia coli	30	1	5.9%	9	24.3%	20	24.1%
Enterobacter cloacae.	2	0	0%	1	2.7%	1	1.2%
Enterococcis fecalis	5	4	23.6%	1	2.7%	0	0%
Hafnia alvei	1	0	0%	0	0%	1	1.2%
Klebsiella pneumoniae	10	1	5.9%	3	8.1%	6	7.2%
Micrococcus luteus	15	3	17.6%	5	13.5%	7	8.5%
Proteus vulgaris.	1	0	0%	0	0%	1	1.2%
Staphylococcus aureus	34	3	17.6%	10	27.1%	21	25.3%
Staphylococcus epidermidis	16	0	0%	3	8.1%	13	15.7%
Staphylococcus saprophyticus	7	0	0%	1	2.7%	6	7.2%
<b>Total</b>	<b>137</b>	<b>17</b>	<b>100%</b>	<b>37</b>	<b>100%</b>	<b>101</b>	<b>100%</b>



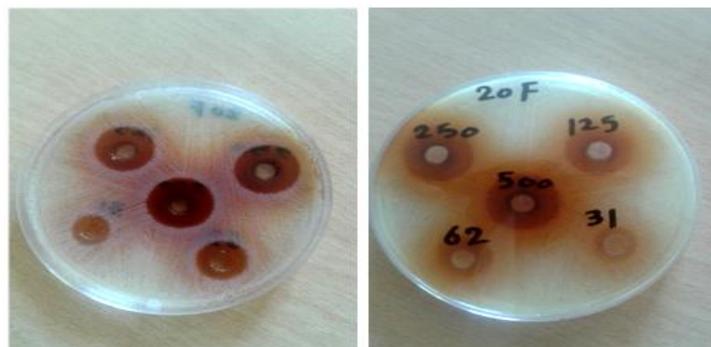
**Fig-1: MIC of aqueous extracts of green tea against tested bacteria (mg/ml)**

**Table-8: Show antimicrobial susceptibility of green tea extract compared to Gentamicin**

Bacteria isolated	Inhibition zone diameter (mm)	
	Aqueous extract (Green tea 500 mg/ml)	Positive control (Gentamicin 10 mcg)
Citrobacter diversus.	9.1 ± 0.9	19.5
Citrobacter freundii	9.1 ± 2.1	19
Escherchia coli.	10.8 ± 3.0	18
Enterobacter cloacae.	8 ± 0	24
Enterococcus faecalis.	17.6 ± 1.9	29
Hafnia alvei.	9 ± 0	20
Klebsiella pneumoniae.	8.6 ± 2.5	23
Micrococcus luteus.	16.5 ± 1.9	28.5
Proteus vulgaris.	8 ± 0	20
Staphylococcus aureus.	18.0 ± 2.4	29.1
Staphylococcus epidermidis.	16.8 ± 2.1	28.9
Staphylococcus saprophyticus.	16.8 ± 2.5	30

**Table-9: Antimicrobial activity of green tea extracts on bacteria isolated**

Bacteria Isolated (n=137)	Inhibition zone (mm)
Gram positive bacteria	17.1 ± 0.6
Gram negative bacteria	8.9 ± 0.9



**Plate-1: Green tea water extract inhibition zones of S.aureus as example at different concentrations (500, 250, 125, 62 and 31 mg/ml)**

## DISCUSSION

Infection of urinary tract is a common problem during pregnancy (Alemu, *et al.*, 2012). The prevalence of UTI among Sudanese pregnant women was 14.0% (Hamdan, *et al.*, 2011). This usually begins in first trimester and peaks during second and third trimester of pregnancy due to a number of factors and physiological changes including urethral dilatation, increased bladder volume (Van Brummen, *et al.*, 2006). The main causative agent of UTI in the study population was *S. aureus* (24.8%), which reported as commonest pathogen among pregnant women, this is in agreement with report from Nigeria (Akinloye, *et al.*, 2006), followed by *E. coli* (21.9%). The green tea water extract had a suppressive effect on the growth of *E. coli* strains isolated from UTIs (Reygaert & Jusufi, 2013). In current study antimicrobial susceptibility of aqueous green tea extract show highest area of inhibitory growth against *Enterococcus faecalis* ( $17.6 \pm 1.9$  mm) and lowest zone of inhibition against *Proteus vulgaris* and *Enterobacter cloacae* ( $8 \pm 0$  mm). The antibacterial activity of tea is due to presence of catechins and polyphenols which disrupt bacterial cell membrane (Kumar, *et al.*, 2012). Ikigai and colleagues reported that tea catechins have less activity on Gram negative bacterial cell membrane due to the fact that Lipopolysaccharide outer membrane of Gram negative is negatively charged (Ikigai, *et al.*, 1993), Kumar and colleagues reported that the daily drinking of green tea can kill *Staphylococcus aureus* also including many other harmful bacteria (Kumar, *et al.*, 2012), Alizadeh and Mohebalian reported that the maximum area of growth suppression was observed against *S. aureus* (A Gram positive organism) and the minimum was against *Pseudomonas* (Alizadeh & Mohebalian, 2016). Many studies have shown that 500 µg of tea polyphenols can suppress the growth of *E. coli*, and  $\geq 5000$  µg are considered bactericidal. This effect is believed to be due to tea polyphenols down regulate the production of proteins such as EF-2 (elongation factor for protein translation); proteins involved in phospholipid, carbon, and energy metabolism; and production of proteins involved in amino acid biosynthesis (Reygaert & Jusufi, 2013). The MIC results of green tea water extract on tested bacteria showing that lowest concentration able to suppress the growth of bacteria is (48.6 mg/ml) appear against *S. aureus* followed by *Staphylococcus saprophyticus* (58 mg/ml), *Enterococcus faecalis* (62.8 mg/ml), *Micrococcus luteus* (66.8 mg/ml) *Staphylococcus epidermidis* (68.2 mg/ml), *Hafnia alvei* (250 mg/ml), *Escherichia coli* (280.2 mg/ml), *Citrobacter freundii* (312.5 mg/ml), *Citrobacter diversus* (325 mg/ml), *Enterobacter cloacae*, *Klebsiella pneumoniae* and *Proteus vulgaris* (500mg/ml). Neyestani and colleagues investigated microbiologic effects of tea extract on some antibiotics against *E. coli* in vitro. They used bacterial control strain ATCC 25920 and tea extracts (green and black tea) of different concentrations (6.25 mg/mL, 12.5 mg/mL, 25 mg/mL,

50 mg/mL, and 100 mg/mL). They use disc diffusion protocol for bacterial sensitivity tests. Green tea at 20 mg/mL concentration inhibited *E. coli* growths completely (Neyestani, *et al.*, 2007) which disagree with the study results. We suggested that green tea that prepared with liquid chromatography show high performance than prepared with dry oven.

## CONCLUSION AND RECOMMENDATIONS

The water based green tea extract has ability to suppress the growth of most bacteria that cause urinary tract infection. The lowest concentration of green tea aqueous extract able to suppress growth of bacteria that cause UTI is 48.6 mg/ml the suppressive activity of green tea aqueous extract is better in Gram positive bacteria than Gram negative according to MIC and zone of inhibition results. Further studies recommended with different ethnic groups.

## ACKNOWLEDGEMENT

The authors appreciate to ethical review committee of the selected health facilities and the pregnant women who consented to taking part in the study and microbiology staff in University of Shendi. Funding for this study was from personal contributions from the authors, no external funding.

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